Corneal Regeneration After Photorefractive Keratectomy: A Review

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Abstract Photorefractive keratectomy (PRK) remodels corneal stroma to compensate refractive errors. The removal of epithelium and the ablation of stroma provoke the disruption of corneal nerves and a release of several peptides from tears, epithelium, stroma and nerves. A myriad of cytokines, growth factors, and matrix metalloproteases participate in the process of corneal wound healing. Their balance will determine if reepithelization and stromal remodeling are appropriate. The final aim is to achieve corneal transparency for restoring corneal function, and a proper visual quality. Therefore, wound-healing response is critical for a successful refractive surgery. Our goal is to provide an overview into how corneal wounding develops following PRK. We will also review the influence of intraoperative application of mitomycin C, bandage contact lenses, anti-inflammatory and other drugs in preventing corneal haze and post-PRK pain.

\textsuperscript{*} The authors have not proprietary or commercial interest in the medical devices that are involved in this manuscript.

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PALABRAS CLAVE Queratectomía fotorreactiva; Córnea; Curación de heridas; Lentes de contacto
The ablation surgery of the corneal surface for the correction of refractive errors began with the development of the excimer laser. The acronym laser means "Light Amplification by the Stimulated Emission of Radiation". Photorefractive keratectomy (PRK), developed by Trokel and colleagues in 1983, uses an excimer laser that emits ultraviolet light of 193 nanometers (nm), a combination of Argon and Fluor (ArF) to remodel the corneal.1 It was not until 1996 when the Food and Drug Administration (FDA) approved PRK as a refractive surgery technique.² In PRK the excimer laser acts on the anterior corneal stroma,³,⁴ producing a stromal remodeling, and, consequently, inducing a change in corneal refraction.⁵,⁶ It corrects mild to moderate myopia, hyperopia and astigmatism, with high level of safety and efficacy.⁷ However, the use of PRK has been reduced over the past years by the introduction of the Laser In Situ Keratomileusis (LASIK).⁸,⁹ Although LASIK provides significant advantages over PRK, such as less surgical trauma, shorter recovery time, and better visual outcomes, it is associated with higher surgical risk, longer recovery time, and a higher incidence of complications.¹⁰,¹¹ In addition to the above-mentioned advantages, LASIK may also be associated with higher rates of complications, such as dry eye, flap complications, and refractive errors.¹²,¹³ It is also indicated in military pilots, professional athletes, or patients that have a high risk for traumatic postoperative flap dislocation.¹⁴,¹⁵ In addition to the long-term advantages and safety, the PRK has gained popularity with the recent wave front guided laser ablation, which reduces postoperative high order aberrations (HOA), improving the optical quality.¹⁶,¹⁷,¹⁸,¹⁹

The visual quality might not be optimal if some complications take place, like subepithelial corneal haze, epithelial hypertrophy, regression of refractive error, deposition of subepithelial extracellular matrix or fibrosis. Other adverse effects include postoperative pain, abnormal corneal nerve regeneration, and night vision symptoms like halos and glare.²⁰,²¹,²²,²³,²⁴,²⁵,²⁶,²⁷,²⁸,²⁹,³⁰,³¹,³²,³³

The purpose of this review is to explain the main cellular changes and complications that occur in different corneal layers after PRK, and to explain how they affect the visual quality. We discuss the role of mitomycin C and bandage contact lenses in corneal regeneration, and the role of different drugs in postoperative corneal pain management.

Corneal Wound Healing

Corneal wound healing is a complex process that, in normal conditions, culminates in the restoration of the tissue, without scar formation or inflammation. The aim is to maintain transparency to recover a proper visual function. After epithelial injury, the corneal healing starts with the removal of necrotic cells. Fibronectin provides a transient matrix for the adhesion of migrating cells, until an epithelial monolayer covers the injured area. Fibronectin also stimulates the production of plasminogen activator (PA), and by a cascade of events, cell-subepithelial matrix adhesions break down. In the next step, limbal stem cells undergo mitosis to reestablish lost cells, and with the anchoring of hemidesmosomes to the underlying stroma, the epithelial regeneration process completes. Stromal wound healing depends on epithelial cells, and on their interaction with keratocytes. Following stromal injury, released cytokines induce the apoptosis of keratocytes under the wound, and stimulate the proliferation and migration of neighboring keratocytes. These active keratocytes synthesize matrix metalloproteinase (MMP) to remodel the stroma. At later stages, a number of them take the repair phenotype, the so-called myofibroblasts, and produce collagen and extracellular matrix (ECM), until the basement membrane prevents the inflow of cytokines in the stroma, and myofibroblast, presumably, commit apoptosis (Fig. 1).

Epithelial Wound Healing Following PRK

The corneal epithelium is formed by superficial, wing and basal cells. In order to facilitate the stromal ablation in PRK, the corneal epithelium is removed. The absence of the epithelium will condition corneal repair. Corneal epithelial cells are the first cells involved in the corneal regeneration process after PRK. Epithelial cells proliferate and migrate from the limbus and the basal epithelial layer to reestablish corneal layers. Corneal regeneration after PRK can be better understood using current, non-invasive, confocal microscopy. It has been used on animals and on humans for corneal cellular structure visualization in real time. Epithelial cells proliferate and migrate from the limbus and the basal epithelial layer to reestablish corneal layers. Histological studies conducted in animals and in humans, have found that corneal epithelium is thicker after PRK, caused by an elongation of the basal epithelial cells and an increased number of superficial cell layers. The corneal flattening in myopic PRK may result in postoperative epithelial thickening due to

es el logro de la transparencia corneal para restablecer la función de la córnea, así como la calidad visual adecuada. Por tanto, la respuesta de regeneración y cicatrización corneal es esencial para el éxito de la cirugía refractiva. Nuestro objetivo es aportar una visión general sobre el modo en que se desarrolla dicho proceso tras la PRK. Revisaremos también la influencia de la aplicación intraoperatoria de mitomicina C, lentes de contacto terapéuticas, y otros fármacos para prevenir el haze y el dolor tras la PRK. © 2014 Spanish General Council of Optometry. Publicado por Elsevier España, S.L.U. Todos los derechos reservados.
the lack of mechanical influences of the upper eyelid that polishes the corneal surface with blinking.\textsuperscript{4} Epithelial hyperplasia in PRK is associated with deep stromal ablation depths and with small ablation zones (4.00–4.5 mm) because there is a marked curvature change in the edges of the ablated area. When ablation zones are large (6.00 mm), they have less demarcated contours, and thus, the change in epithelial thickness is minimal.\textsuperscript{51–56} Table 1 shows the variation of central corneal thickness with different surgery techniques published in the scientific literature. Erle\textsuperscript{2} proved that, after PRK, the central epithelial thickness returned to preoperative levels at 1 month. However, it continued to progressively increase during the first year, being 21% thicker at that time. This result is similar to the 22% thickness increase seen in LASIK by Erle et al.\textsuperscript{57} However, the time required for thickness stabilization differs between the two techniques, due to the complex interaction of epithelial cells and activated keratocytes in PRK.\textsuperscript{2} According to Patel et al.,\textsuperscript{25} central corneal epithelium in LASIK increased 24% during the first year after surgery and remained stable during the next 7 years. In PRK, corneal thickness continued to increase at 1 month, 1 year, and 7 years (442±39 \textmu m, 464±44 \textmu m, 471±45 \textmu m, respectively).\textsuperscript{25} Recently, Iversen et al.\textsuperscript{58} have concluded that in PRK and LASIK, the epithelial thickness increases 15%–20% after surgery, but the epithelial changes in LASIK occur during the first week and remain unaltered during the following 3 years. It has been suggested that epithelial hyperplasia can induce a reduction of postoperative refractive effect. Erle showed myopic regression significantly associated with epithelial thickness increase.\textsuperscript{2} Nevertheless, Iversen et al.\textsuperscript{58} did not find any correlation between changes in epithelial thickness and changes in refraction after PRK or LASIK, probably because of the small size of their sample.

**Stromal Wound Healing Following PRK**

Stroma occupies approximately the 90% of corneal thickness,\textsuperscript{59} and it can be subdivided into three continuous layers: anterior, middle and posterior.\textsuperscript{59} The corneal stroma is built up from collagen fibers, ground substance, keratocytes and nerve fibers.\textsuperscript{5,49} Keratocytes – corneal stromal cells – play a major role in maintaining corneal transparency, and synthesizing the components of the extracellular matrix (ECM).\textsuperscript{56} Active keratocytes produce collagen and proteoglycans to form the ECM after stromal injury. The human stromal cornea contains collagen type I, V and VI.\textsuperscript{39,60} Type I is predominant (75%), followed by type VI (approximately, 17%).\textsuperscript{60} Type III collagen appears in inflammatory events or during wound healing. Proteoglycans participate in collagen fibrillogenesis and matrix assembly.\textsuperscript{44} After corneal injury, newly produced collagen fibers tend to have larger diameters, as they contain high levels of dermal sulphate (a type of proteoglycan) that lasts up to 6 months.\textsuperscript{59}

Stromal keratocytes are normally quiescent or inactive, and are the second cells involved in the process of corneal regeneration, just after corneal epithelial cells. After PRK, keratocytes underlying the wound disappear by apoptosis due to a stress exposure,\textsuperscript{2,24,37,40,30,63} During the first 24h after injury, macrophages, monocytes, T cells and polymorphonuclear cells infiltrate the area and remove damaged cells.\textsuperscript{41,64} Metalionoproteinases (MMPs) and the plasminogen activator system remove the affected extracellular matrix.\textsuperscript{9,65–67} The MMPs are proteolytic enzymes secreted by active keratocytes or fibroblasts, and degrade complex molecules of the extracellular matrix. Although nine types of MMPs exist, in the cornea only four MMPs are important, being MMP-1 the most relevant.\textsuperscript{67} MMP-8 concentration has been observed to be significantly elevated in the second day after PRK (P=0.001).\textsuperscript{68} The remaining keratocytes, adjacent to wound borders, are activated in response to various cytokines released by cells in upper layers, such as interleukin (IL)-1, and growth factors like tumor necrosis factor (TNF), fibroblast growth factor (FGF), platelet-derived growth factor (PDGF), epithelial growth factor (EGF), and transforming growth factor (TGF).\textsuperscript{2,8,14,22,24,35,51,58,69–74} These growth factors are normal components of the tear and corneal cells, produced by the lacrimal gland,\textsuperscript{15} and regulate a variety of processes involved in homeostasis and corneal wound healing, including migration, mitosis and cell differentiation.\textsuperscript{75} Particularly, transformation growth factor beta (TGF-beta) seems to transform active keratocytes into myofibroblasts that appear at later stages of stromal heal-
PRK produces oxygen free radicals, secondary to the exposure of ultraviolet radiation, thermal increase, and polymorphonuclear cell infiltration. Free oxygen radicals may interact with lipid components, nucleotides, and sulphur contained in enzymes, and particularly with reactive oxygen species (ROS) that are considered to produce the most reactive and cytotoxic damage. In fact, they have been described as a partial cause of keratocyte apoptosis. Among the antioxidant enzymes that protect the cornea from radicals, superoxide dismutase (SOD), glutathione peroxidase (Gpx) and catalase are the most relevant. Ascorbic acid and α-tocopherol (Vitamin E) also prevents from the effects of free radicals.

Corneal epithelial ascorbic acid absorbs ultraviolet radiation, protecting keratocytes, but high or altered corneal ascorbate levels in the human cornea after PRK, may produce accelerated keratocyte death. In rabbit corneas, decreased activity of SOD and Gpx enzymes has been proved after refractive surgery. For this reason, additional antioxidant enzymes seem to be involved in reducing corneal oxidative stress following PRK. 1-cys peroxiredoxin (1-cys Prx) may be an important enzyme involved in the differentiation, migration and proliferation of epithelial cells. 1-cysPrx increases 4 h after PRK and remains in high levels until 7 days after PRK.

The density of keratocytes varies across the stroma. It is estimated that in the anterior stroma the density is 5%–10% greater than in middle and posterior stroma. It has been documented that a corneal stroma rich in keratocytes prevents the epithelial corneal infection or, at least, minimizes the extension of the infection. After PRK, the anterior keratocyte population drastically diminishes, and the distribution and shape is greatly altered. In confocal microscopy, high reflectance, hyperplasticity, hypertrophy and a decrease in the contrast of the anterior stromal keratocytes can be observed. Human histological studies confirm that the decrease of anterior stromal keratocytes in humans and animal respond similarly. Table 2 shows the variation of anterior, posterior and total keratocyte density in the different studies published in the scientific literature. It can be seen that after 5 years of PRK there were evidences of keratocyte density loss in middle and posterior stroma. They observed a reduction of 20%–24% in the posterior stroma (P < .05) although they claimed that this loss was not completely evident. Keratocyte density in the anterior 10% of the stroma continues to decrease 5% per year between 1 and 3 years after PRK. Erin et al. reported a progressive decline in anterior stromal keratocytes, becoming significant at 36 months after PRK (P = .02). In contrast, middle and posterior keratocyte densities remained unchanged between 1 and 3 years after PRK. In another study, Erin et al. proved that the keratocyte density in the anterior 10% of the stroma, decreased at 6, 12, 24 and 36 months (41%, 40%, 43%, 45%; respectively) after PRK, compared to pre-PRK. In a posterior longer-term study, Erin et al. demonstrated a similar decreasing pace in anterior keratocyte density: 40%, 42%, 45%, and 47% at 6 months, 2 years, 3 years, and 5 years (P < .001). Amoozadeh et al. found a reduction in keratocyte density 6 months after surgery, but the loss was similar for LASIK and PRK interventions: in

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<th>Study</th>
<th>Technique</th>
<th>Preop</th>
<th>1 month</th>
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Table 1: Variation of Central Corneal Thickness, Mean±SD or Range (μm).

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Table 2: Keratocyte Density in the Anterior, Posterior and Total Stroma in μm²/mm².
anterior stroma, 34.7% versus 31.13% (P > .05) and posterior stroma 0.31% versus 0.02%, (P > .05), respectively. However, other studies have seen differences between PRK and LASIK, probably associated with the more superficial ablation in PRK. The consequences of keratocyte density loss after PRK are still unknown, but the visual acuity and corneal clarity seem to be preserved.

After the initial depletion of anterior stromal keratocytes, an increase in the keratocyte density is observed over time, probably secondary to mitosis, cellular migration, or reproduction of keratocytes and myofibroblasts. Following apoptotic keratocyte loss, the first morphological changes of remaining keratocytes that can be histologically observed, are an increase in cell size and an increase in the size and the number of nucleoli, rough endoplasmatic reticula, mitochondria, free ribosomes and Golgi complexes, indicating an active state. These keratocytes quickly repopulate the anterior stroma, and return to similar preoperative levels. Several studies using confocal microscopy have analyzed the keratocyte density after PRK. Corbett et al. found that at 2 days after PRK the anterior keratocyte density was increased 50%, 100% at 1 month, and returned to preoperative levels at 6 months. Frueh et al. concluded that the anterior keratocyte density increased 15% at 1 and 4 months after PRK, and returned to preoperative levels 1 year after PRK. Similarly, Erie et al. found an increase of 20% in the anterior stroma at 3 months after PRK. According to the results of Corbett et al. and Erie et al., anterior keratocytes proliferation begins 1 month after PRK, with a pick at 3 months, and return to preoperative levels at 6 months.

**Corneal Haze**

Corneal haze reduces corneal transparency at variable degrees. Subepithelial haze occurs in all patients 1 month after PRK, reaching the greatest intensity at 3–6 months, and gradually decreases from then on. Yet, some authors affirm that it begins to decrease at 12–24 months after PRK. Corneal haze is more common after correction of high myopia (>-6.00 D), and it is rarely seen after correction of <-6.00 D of myopia or <+4.00 D of hyperopia. Besides the ablation depth, the severity of corneal haze is correlated with excessive ocular UV-B radiation, duration of the epithelial defect, postoperative steroid treatment, male sex and with certain population with brown irises. In the presence of damaged epithelial cells and basement membrane, cytokines and growth factors can easily flow from epithelium to anterior stroma. Cytokines released from epithelial cells activate keratocytes, as mentioned in a previous section, which synthesize large diameter collagen fibrils. Abnormally deposited extracellular matrix implies the development of corneal opacity. Moreover, active keratocytes present a high reflectance that also contributes to the decrease in corneal transparency. In addition, subepithelial vacuolation, deposit materials like proteoglycans, hyaluronic acid and collagen Type IV are involved in the formation of the corneal haze in advanced stages. Plasminogen activator–plasmin
system degrades the damaged ECM, and extended low levels beyond the third day after PRK causes corneal haze formation. Guerrero et al. affirm that the loss of collagen type IV is related to the activation of keratocytes in vivo and in vitro, and Winkler et al. and Mohrenfels et al. emphasize on the role of type IV collagen in the development of corneal cloudiness. Secondary ultraviolet B (UV-B) exposure, originating from sun or solarium is a causal factor for aforementioned abnormal proteoglycan deposition and associated augmented corneal thickness.

On the other hand, myofibroblasts, derivatives of TGF-beta responding keratocytes, are thought to be the first biological event for corneal haze formation. Myofibroblasts play an essential role in the recovery of the corneal integrity after penetrating injury, mainly in advanced stages. They secrete extracellular matrix, contract wounds and have the ability to generate adhesion structures with the surrounding substrate. TGF-beta also induces the expression of connective tissue growth factor (CTGF), which mediates collagen synthesis, and along with myofibroblasts regulates the corneal wound healing, and may promote scar formation. After PRK, myofibroblasts appear as a pathological response to injury, and their decreased transparency roots in the low intracellular content of crystalline. Irregular surface has also been related to high incidence of corneal haze, and higher irregularity is seen with increasing dioptric corrections in PRK. Interestingly, surface irregularity is positively correlated with myofibroblast density in the anterior stroma. In normal corneal wound healing, complete regeneration of the basal membrane after PRK occurs within 6–8 weeks in rabbits, which limits the access of growth factors to the stroma and, consequently, myofibroblasts commit apoptosis modulated by IL-1. Therefore, the presence of myofibroblast, and subsequent corneal haze, is largely dependent upon the restoration of the basement membrane.

Corneal haze has been traditionally measured in the slit-lamp, and graded with diverse scales, like Hanna’s scale. The new technology leads us to use automated instruments for corneal haze measurement. In vivo confocal microscopy is a reliable tool, as far as standardized methods are used. It is the most widely used objective method in clinical setting for haze measurement. In the last years, alternative techniques have come out. Confocal imaging of second harmonic-generated (SHG) signals has been shown to be sensitive in measuring corneal fibrosis after refractive surgery. Recently, the densiometry program of Pentacam Scheimpflug imaging system (Oculus Optikgeräte GmbH) has been proved to be a useful method for measuring corneal haze.

Visual Disturbances of Corneal Haze

The corneal haze produces a reduction of low contrast visual acuity and night vision symptoms that, in the vast majority of situations, improve with time. It is possible to see corneal haze formation after PRK by means of confocal microscopy, observed as a decrease in the contrast of the image and an increase in reflectivity. Böhne et al. using a Tandem scanning confocal microscopy, correlated corneal haze and anterior stromal reflectivity. However, the tandem scanning confocal microscopy is not able to detect acellular regions of the anterior stroma early after PRK when epithelium and sub-basal plexus are not formed. Although corneal haze in humans is less pronounced than in animal models, if corneal haze persists and affects significantly to the corneal transparency, it causes light scatter and secondary visual disturbances. For this reason, corneal haze may be described and analyzed through back light scattering (backscatter). It also causes irregular astigmatism and subsequent loss of corrected distance visual acuity (CDVA).

The regression of the refractive error may be produced by epithelial irregularity, alterations in the keratocyte density or subepithelial deposits. Myopic regression occurs in 78% of eyes in the first 12 months after PRK. Table 3 shows the mean spherical equivalent changes reported in different scientific studies. In the first week after PRK, epithelial irregularity causes a reduction in visual quality. During the first month, altered keratocytes decrease contrast sensitivity, mainly in high frequencies, and cause glare. During the next 2 months, subepithelial deposits produce a decrease in contrast sensitivity, especially in low frequencies. Ginis et al. reported that subepithelial deposits are the first factor that contributes to the development of corneal scatter. The visual quality is affected temporarily, although there is evidence that in some cases it persists for more than 1 year. In order to avoid a decrease in the visual quality, all postoperative efforts must go oriented to control the subepithelial matter. The corneal epithelium does not seem to contribute significantly to the refractive change after PRK, although some studies suggest that epithelial thickening may produce myopic regression, even 5 years after PRK. Moller-Pedersen et al. and Cua and Pepose suggested that new keratocytes growth in central cornea or postoperative corneal scarring is likely to be the main causes of myopic regression in ablations of 6 mm. In agreement with this hypothesis, Moller-Pedersen et al. demonstrated that hyperopic changes were the direct result of a stromal thinning. Erie found an increase of 12 μm of epithelial thickness at 12 months after PRK that was associated with a myopic regression of −0.41 diopters but no correlation was found between stromal thickening and myopic regression; however, the combined effect of epithelial and stromal thickening was correlated with myopic regression.

Regeneration of Corneal Innervation

The cornea is the most innervated tissue of the human body, and these sensory nerves are derived from the trigeminal branch of the trigeminal nerve fibers. Corneal sensory nerves penetrate the limbus and form nerve bundles in the anterior third of the stroma. Once there, they run perpendicularly to cross Bowman’s membrane, and form the sub-basal nerve plexus as a network between the basal epithelial cells and Bowman’s layer (Fig. 2). Corneal nerve fibers, if visualized using confocal microscopy in normal conditions, show high reflectivity across the corneal stroma with a rectilinear pattern. Subepithelial nerve fibers, on the other hand, are thinner than stromal nerve fibers. Corneal fibers are considered primarily nociceptive (70%), followed by mechanosensitive fibers (20%). In PRK, ph-
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<th>Study</th>
<th>Technique</th>
<th>Preop</th>
<th>1 month</th>
<th>6 months</th>
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<td>PRK+AAD</td>
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<td>(−4.00 to −1.25)</td>
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<td>Wallau and Campos (2008)</td>
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<td>(−1.46 to −6.96)</td>
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<td>PRK+AAD</td>
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<td>(−4.00 to −1.25)</td>
<td>−0.28±0.91</td>
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<td></td>
<td>PRK+MMC</td>
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<td>(−1.95 to −6.40)</td>
<td>0.61±0.61</td>
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<td>Ghirlando et al. (2007)</td>
<td>PRK</td>
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<td>−0.08±0.38</td>
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<td>−0.2±0.5</td>
<td>(−1.25 to −0.75)</td>
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<td>PRK</td>
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<td></td>
<td>−0.3±0.3</td>
<td>(−0.87 to plano)</td>
<td>−0.4±0.4</td>
<td>(−1.25 to plano)</td>
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<tr>
<td></td>
<td>LASEK</td>
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<td>0.24±0.61</td>
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<td>−0.16 to −1.10</td>
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</table>

PRK, photorefractive keratectomy; MD, mechanical debridement; AAD, alcohol-assisted debridement; LASIK, laser in situ keratomileusis; MMC, mitomycin C; LASEK, laser-assisted subepithelial keratectomy; tPRK, trans-PRK.

* (0.002%, 1 min).

* (0.02%, 2 min).
Substances According

However, using conversely, at 8 weeks after PRK, sub-epithelial nerve fibers are visible on the edges. Erie, using tandem scanning confocal microscopy, visualized subbasal nerve fiber bundles in 17% of the corneas at 1 month after PRK. However, he noted that the density of these nerve fibers was 98% less than preoperatively. After about 3 months of the surgery, no branched nerve fibers can be visualized in the center of the zone of ablation. Changes in subepithelial plexus and stromal trunks begin to appear 2–4 months postoperatively. At 6–8 months after the intervention of PRK subepithelial nerve regeneration is almost complete, although changes in the structure of the corneal nerves can be appreciated by confocal microscopy up to 12 months postoperatively. However, nerve density continues to improve until 12 months after surgery, and returns to the preoperative values at 2 years. According to Moilanen et al. in 71% of cases the central branching postoperatively was comparable to control subjects at 5 years (P=.56). Erie proved that subbasal nerve density was reduced at 3, 6 and 12 months (87%, 75%, 60%, respectively) after PRK, and returned to preoperative levels at 24 and 36 months postoperatively. Subsequently, Erie et al. in a prospective 5-year longitudinal clinical trial, proved with confocal microscopy that the recovery of subbasal nerve density in central cornea was faster in PRK than in LASIK. The authors observed that subbasal corneal density was reduced by 59% at 1 year after PRK (2764±1321 μm/mm²) compared to preoperatively (6786±1948 μm/mm²; P<.001). Sub-epithelial plexus was almost recovered 2 years after PRK (6242±1763 μm/mm²), and remained unchanged at 3 years (6358±2447 μm/mm²) and 5 years (5903±3086 μm/mm²). In LASIK, they observed that subbasal corneal density was reduced by 34% at 3 years (P<.001) with values at 5 years postoperatively comparable to those obtained preoperatively (5903±3086 μm/mm²). It is worth to note that in this study, the corneal flap was created using a mechanical microkeratome. As the new technology allows making corneal flaps with laser instead of with a mechanical microkeratome, it is possible that studies in the near future report a faster corneal nerve recovery after LASIK.

When the process of corneal nerve regeneration finalizes, morphological abnormalities are often observed. According to Erie, in the first 6 months after PRK the central subbasal nerves are organized in horizontal or oblique orientation. However, between 6 and 12 months, the subbasal nerve orientation rotes and comes to vertical orientation. In dry eye conditions, Esquenazi et al. observed active keratocytes, and they expressed nerve growth factor (NGF). NGF stimulates the proliferation of basal epithelial cells in normal conditions. Active keratocytes provoke an overexpression of NGF, which leads to abnormal findings in corneal nerves, such as hypertonphy. They also found higher nerve tortuosity, higher number of nerve beads, and the presence of nerve sprouts in desiccating environment group, which
Corneal Regeneration After Photorefractive Keratectomy

means there is a high metabolic activity to repair the alterations in the corneal epithelium.

**Corneal Pain and Sensitivity**

Photoablation severs corneal nerves, disrupting the lacrimal functional unit (LFU). LFU is constituted by the lacrymal gland, ocular surface and innervation. It regulates tear secretion, and affects its composition.123 Thereby, photoablation produces transitory dry eye, deterioration of corneal barrier function and alteration in corneal sensitivity.83,111,124 A reduction of the tear flow after PRK has been proved using Schirmer test.16 According to Erie et al.,83 LASIK presents higher prevalence of postoperative dry eye, altered corneal epithelium and tear film than PRK. Dry eye has been associated with low corneal sensitivity.125,126 Different devices are available to measure corneal sensitivity, as Cochet-Bonnet esthesiometry or non-contact gas esthesiometer. Cochet-Bonnet esthesiometer only stimulates mechanosensory fibers, whereas non-contact gas esthesiometer measures activation thresholds of nociceptors using controlled chemical, thermal and mechanical pulses. Non-contact gas esthesiometer is, therefore, a more sensitive device for measuring alterations in corneal sensitivity. Still, Cochet-Bonnet esthesiometry is more widely used, and controversy remains about the time course of the corneal sensitivity recovery after PRK with this device. Kauffmann et al.130 affirm that the recovery of corneal sensitivity usually starts at 4–6 weeks, completing approximately within 6–12 months following PRK. However, Erie et al.83 claim that the recovery of corneal sensitivity is completed from 3 months to 1 year after PRK. Hypoesthesia is often expected until 3 months after surgery, due to the loss of corneal nerves.8 However, the other hand, Gallar et al.127 measured corneal mechanical and chemical sensitivity following PRK with non-contact gas esthesiometer, and found that both types of sensitivities were reduced even 5 years postoperatively, achieving normal values in 10 years. Despite the diminished corneal sensitivity, intense pain is usually present hours after PRK.128 Gallar et al.129 attributed corneal pain and discomfort sensations to the altered functionality of corneal nerves. They recorded spontaneous activity and modified responsiveness in corneal fibers of cats that underwent PRK.129 Experimental evidences support the idea that ongoing activity evokes spontaneous pain sensations.130,131

**Acceleration of Corneal Regeneration Process, Reduction of Corneal Haze and Corneal Pain Management**

Nowadays there are several alternatives to speed up the process of epithelial regeneration, like epithelial removal techniques, amniotic membrane, or bandage contact lenses. In PRK, agents like mytomycin-C (MMC) or fluoroquinones that reduce the corneal haze formation are used, and drugs to reduce the corneal pain and inflammation are also prescribed.

**Epithelial Removal Techniques**

In PRK, previous to the impact of laser energy over the cornea, the corneal epithelium has to be removed. The removal of the corneal epithelium is carried out mainly with epithelial mechanical scraping using chemical agents like diluted ethanol solution, through a rotary brush or using the laser itself – known as transepithelial ablation (Fig. 3).18,21,22,29,61,128,132,133 The epithelial scraping has postoperative adverse effects like pain, myopic regression or corneal haze. Some modification in PRK technique can alter the wound healing response with the aim of minimizing the adverse effects.23 The exposition to agents such as ethanol can produce an increase in the inflammatory response and more damage to the anterior stromal keratocytes that could increase the haze formation.21,24 Yet, controversy remains in the scientific literature because other authors affirm that alcohol-assisted epithelial removal produces less inflammation, favoring epithelial regeneration and preventing corneal haze or keratoocyte apoptosis.9,62 Esquenazi et al.22 proved that the epithelial scraping might be associated with an increase in the number of reflective structures in the stroma, mainly in cornes with ocular dryness after PRK. The laser-scare epithelial removal decreases the degree of keratoocyte apoptosis, producing a less pronounced loss of superficial keratocytes.2 However, the irradiation with cold balanced salt solution (BSS) may alter the keratoocyte apoptosis in the retroablation zone. The time necessary for mechanical debridement is greater than the time required for laser or alcohol scrape techniques, even for expert surgeons.18 Mechanical debridement is related to stromal dehydration and disappearance of anterior stromal keratocytes.6,63 This loss provokes an increase of cells in the underlying stroma, causing stromal hyperplasia and haze formation.18 Einollahi et al.63 found faster mean epithelial healing time in the alcohol-assisted group than in the mechanical group (3.0±0.3 versus 3.2±0.4 days, P=0.001). They observed greater anterior retroablation stromal keratocyte density in the mechanical group than in the alcohol-assisted groups at 3 months (704.3±119.9 cells/mm² versus 743.3±103.7 cells/mm², P=0.05) and at 6 months (643.8±134.4 cells/mm² versus 696.7±129.6 cells/mm², P=0.02). In the same study, Bahram et al. did not found statistically significant differences in middle and posterior keratoocyte density between the mechanical and alcohol-assisted groups.62 They also proved that mechanical and alcohol-assisted epithelial debridement after PRK present similar visual and refractive outcomes in patients with mild myopia,63 in agreement with the results of Goreishi et al.135 They reported similar safety and efficacy with alcohol-assisted and mechanical debridement in a 1250 eye sample, but anterior keratoocyte density was not assessed in this study.136

Laser-assisted subepithelial keratomileusis (LASEK) was developed in order to reduce corneal pain and haze formation associated with PRK, and to accelerate visual recovery. Epithelial delamination with diluted alcohol showed in an electron microscope study that was able to leave a smooth surface, ideal for LASEK intervention.136 It seems that a regular surface before laser application helps corneal healing and prevents haze.137 Chen et al.138 contrasted these findings in a later study, and showed a high variability.
in morphological changes after diluted alcohol treatment, dependent upon concentration and time. Cell viability was affected when alcohol exceeded its concentration by 25% or 25-s exposure.\(^\text{138}\) Yet, these studies have been conducted in vitro, and the complex interactions of tear film and corneal surface were not considered. In vivo studies do not show any difference between LASEK and PRK.\(^\text{139}\) Lee et al.\(^\text{18}\) evaluated epithelial healing, postoperative pain and visual outcomes using epithelial mechanical (conventional PRK), transepithelial PRK and 20% diluted alcohol laser-assisted subepithelial keratomileusis (LASEK) with flap repositioning. After 6 months, they found little differences in clinical outcomes between the 3 techniques, noting a slight overcorrection in the transepithelial PRK and slight undercorrection in LASEK. Corneal pain and subepithelial haze results were similar.\(^\text{18}\) Subsequently, Ghanem et al.\(^\text{139}\) proved in a prospective randomized double-masked study that the reepithelialization was faster in a PRK group compared with a butterfly LASEK group, even though epithelial semi-discs were repositioned intraoperatively in LASEK group. (4.35±0.48 days versus 4.75±0.72 days, \(P=.002\)). They also found lower pain level in PRK group, but pain scores and ocular discomfort were not statistically different from butterfly LASEK (3.31±4.09 versus 4.43±4.27; \(P=.18\)).\(^\text{139}\)

It has been proven in animal studies that transepithelial ablation produces a uniform surface for corneal regeneration, and prevents kerocyte apoptosis,\(^\text{36}\) reducing the risk of corneal haze.\(^\text{21,72}\) Wang et al.\(^\text{140}\) presented promising preliminary results of SCHWIND-ESIRIS excimer laser for transepithelial ablation, but the flawed design of the study makes difficult to assess the real value of this technique. Later, Aslanides et al.\(^\text{21}\) proved in humans that transepithelial ablation was safer than the epithelial mechanical scraping using chemical agents as alcohol, as it provides a faster epithelial healing, less postoperative pain and less corneal haze at 1 week (\(P=.07\)), and at 1, 3, and 6 months after surgery (\(P<.05\)). In addition, they observed an improvement of 3 Snellen lines in visual acuity on day 3 in the modified transepithelial PRK (all-surface laser ablation) group compared to conventional alcohol-assisted PRK group (0.4 versus 0.2; \(P<.05\)).\(^\text{21}\) Trans-epithelial ablation also resulted in better corrected distance visual acuity (DCVA) than conventional alcohol-assisted PRK (33% versus 13%, respectively, \(P<.05\)).\(^\text{21}\) although differences in higher order aberrations were not statistically significant.\(^\text{21}\)

**Amniotic Membrane Transplantation**

Apart from the above mentioned techniques, amniotic membrane transplantation reduces the inflammation after PRK, prevents polymorphonuclear cell infiltration, produces less peroxidation, avoids kerocyte apoptosis and stimulates corneal epithelialization.\(^\text{137,77}\) It is usually combined with PRK to treat corneal dystrophies, corneal degenerations, scars, keratopathies,\(^\text{141,142}\) or even to treat corneal haze secondary to PRK.\(^\text{143}\) The amniotic membrane restricts the influx of polymorphonuclear cells (PMCs) to the patch.\(^\text{144,145}\) PMCs adhere to the amniotic membrane and eventually commit apoptosis.\(^\text{146}\) This is a physiological way of suppressing corneal inflammation.\(^\text{147}\) In addition, amniotic membrane has intrinsic kerocyte growth factors, EGF and neurotrophins that promote epithelization.\(^\text{148,149}\) It also suppresses TGF-beta1, collagen III and fibronectin.\(^\text{150}\) Taken together, amniotic membrane has a potent anti-scarring effect that reduces corneal haze formation, as demonstrated in animal studies.\(^\text{144,151}\)
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Agents to Enhance Wound Healing

The wound healing response may be altered by the prophylactic application of a topical solution of mitomycin-C (MMC) immediately after the laser ablation, in order to avoid or minimize myofibroblast activation. Mitomycin-C is an antineoplastic antibiotic agent of the family of anti-tumor quinolones and derived from Streptomyces caesipitosus. It is a potent DNA crosslinker: it inhibits the replication of deoxyribonucleic acid (DNA) in the cell nucleus. Wallau and Campos obtained better UCVA and BSCVA with the combination of PRK with MMC, than with LASIK (P<.027 and P<.001, respectively) at 3, 6 and 12 months after surgery. Goreishi et al. reported an incidence of 4% of corneal haze at 1 year postoperatively with intraoperative application of 0.02% MMC, in a sample with a mean refraction error of –5D. Fazel et al. found that two-step administration of 0.02% MMC (45 s, followed by 15 s) further decreased corneal haze formation in high myopia, compared to a single dose of 45 s. The benefits of MMC have also been described since the haze has been established, where mechanical epithelial scraping and instillation of MMC restores corneal transparency.

Although the application of the mitomycin C is helpful for corneal recovery, it is necessary to control the doses and the time of exposure. According to Thornton et al., the concentration is more important than the duration of MMC exposure in corneal haze prevention. Rajan et al. analyzed the effects of MMC after correction of −9.00 diopters by PRK in 3 groups of healthy rabbits: with MMC application, with MMC (0.2 mg/ml) application for 1 min and with MMC (0.2 mg/ml) application for 2 min. The 2 min MMC group (0.2 mg/ml) had thinner epithelium than the 1 min and without MMC application groups (P<.0001). The application of the intraoperative MMC lasts between 10 s and 120 s, depending on the surgeon. According to Khoury et al., the application of intraoperative MMC vary from 12 s to 5 min. Shojaei et al. affirm that short-time MMC exposure prevents low-grade haze in low ablation depths. The MMC doses oscillate between 0.002% and 0.06%. The intraoperative application of 0.02% MMC solution is the most recommended, as it produces less corneal haze, and provides better uncorrected visual acuity (UCVA) and best spectacle-corrected visual acuity (BSCVA). Still, Ramjoo et al. found similar refraactive and haze outcomes with 0.01% and 0.02% MMC for mild myopia, recommending the use of 0.01%. The lowest dose available is recommended to avoid side effects. Rajan et al. observed a delay in keratocyte regeneration after MMC application (P<.0005). Midena et al. proved by means of confocal microscopy that the application of 0.02% MMC produced a considerable decrease of anterior stromal keratocytes, but there is no evidence of this decline in the posterior stromal keratocytes. Subsequently, Thornton et al. observed a keratocyte loss in the anterior stroma 1 month and 6 months after PRK with standard MMC concentrations (0.02%). Razmjoo et al. did not found significant reduction in keratocyte density after application of 0.02% mitomycin C (MMC). The dose of MMC is associated with the grade of refractive error. Thornton et al. believe that for high myopia corrections (≥−6.00 D) standard concentration of topical MMC (0.02%) may be used, whereas for moderate myopia (−3.00 to −5.90 D) low dose of MMC (0.002%) may be considered, although it seems that intermediate dose of MMC (0.02%) is more effective than 0.002% for moderate myopia.

The cytotoxicity of MMC increases with cumulative doses, and when MMC is combined with ethanol, which increments the apoptosis of keratocytes. Few complications have been associated with its use with the exception of a decrease in the short term of the keratocyte density. There are some complications have been documented at the time of instillation or after some weeks. Although unusual, sceral ulceration, non-healing conjunctivas and complications associated with high MMC doses (0.04%) or prolonged postoperative topical use may appear, because high doses of MMC suppress cellular RNA replication and protein synthesis. As MMC is applied in the stromal bed, it seems that it might penetrate into the anterior chamber, because cytotoxic effects on the ciliary body epithelium have been reported. There is controversy in the scientific literature, but MMC does not seem to cause any alteration in the ciliary body or intraocular pressure (IOP) after PRK. Kymionis et al. investigated the effects of MMC after PRK in 40 eyes of 20 rabbits. They applied 0.02% MMC for 2 min in one eye, and balanced salt solution (BSS) for 2 min in the contralateral eye. After 3 months, they did not found differences in the morphology of the ciliary body, and tonometric measurements remained stable (P<.075).

The endothelium is the inner layer of the cornea. Endothelial cells have a hexagonal or polygonal shape, and they are homogeneously distributed, without signs of polymegetasimis and pleomorphism in normal conditions. Endothelial cells are not able to regenerate, and a reduction in the number of cells is seen with age. After PRK, endothelial structure, shape and density remain unaltered. Table 4 shows the variation of endothelial cells in the different studies published in the scientific literature. Polymegetasim or pleomorphism, if present, may be secondary to still unknown corneal metabolism. There is also controversy about the toxic effect of MMC in the overall morphology of the endothelium. Morales et al. proved that intraoperative 0.02% MMC during 30 s after PRK induced corneal endothelial cell loss at 1 month and 3 months (P<.0006, P<.002; respectively). Diakonis et al. applied Mitomycin C (MMC) for 15 s and the density of endothelial cells was not affected. Zare et al. obtained similar results when 0.02% MMC was applied for 45 s. Subsequently, Shojaei et al. found significant differences of mean endothelial cell densities in the MMC group and in the control group at 6 months after surgery (2878.79±283.04 cells/mm² versus 2826.19±286.25 cells/mm², P=.25). Undoubtedly, after the application of MMC the DNA of endothelial cells gets damaged. It remains to be determined the long-term effects of such event. According to Wilson, long-term studies (more than 10 years) are necessary to determine the adverse effects of MMC.
Table 4 Variation of Endothelial cell density After Surgery, Mean±SD or Range (cell/mm²).

<table>
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<tr>
<th>Study</th>
<th>Technique</th>
<th>Preoperatively</th>
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<th>3 months</th>
<th>6 months</th>
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<td>PRK with BSS</td>
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<td>2826±286</td>
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<td>PRK with MD</td>
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<td>Epi-LASIKb</td>
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<td>PRK+MMCd</td>
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<td>Morales et al. (2006)</td>
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<td>2779±492</td>
<td>2711±555</td>
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PRK, photorefractive kerectomy; MMC, mitomycin C; BSS, balanced saline solution; MD, mechanical debridement; AAD, alcohol-assisted debridement; LASIK, laser in situ keratomileusis; Epi-LASIK, epipolisor laser in situ keratomileusis.

a After radial keratotomy.
b 0.002%, 1 min.
c 0.02%, 2 min.
d 0.02%, 30 s.
e 0.02%, 15 s.

New generation quinolones, instead of preventing corneal haze, are used as prophylactic antibiotics to avoid corneal infections after refractive surgery. They also enhance the rate of corneal recovery. Fourth generation fluoroquinolones like gatifloxacin (Zymar, Allergan, Irvine, California) and moxifloxacin (Vigamox, Alcon Laboratories, Fort Worth, Texas) have been demonstrated to mediate faster corneal healing, without evident differences between both of them in terms of visual outcomes.

Bandage Contact Lenses

After PRK, the corneal surface needs between 2 and 4 days to regenerate, and the vision may fluctuate for several weeks to months. If epithelial regeneration delays, the subepithelial haze increases; for this reason, an appropriate corneal reepithelialization is crucial. Reepithelialization is the first step during corneal regeneration after PRK. If the reepithelialization is facilitated with the appropriate contact lenses, visual acuity improves. Although therapeutic contact lenses have been used for more than 40 years, PRK has increased their popularity. One of the major disadvantages of PRK is the pain and discomfort during 1–3 days after intervention. To ease off the postoperative pain and discomfort, and to promote epithelial healing, bandage contact lenses are fitted for 3–5 days after surgery. Other techniques and medications have been proposed in order to reduce corneal pain like occlusive pressure patching, but the bandage contact lenses are still the gold standard. Bandage contact lenses are used to protect the epithelium from the eyelid, to reduce the haze formation, to enhance epithelial healing, to control the sensation of pain, and to prevent epithelial erosions. Faster reepithelialization produces a reduction of discomfort, facilitates visual recovery, and restores the corneal barrier to prevent infections. Because of the prolonged use of therapeutic contact lenses, and to assure the proper corneal metabolism, a high oxygen permeability (Dk/t) contact lens are used. Silicone hydrogel contact lenses have a Dk/t coefficient 5–10-fold greater than conventional hydrogel lenses. For this reason, silicone hydrogel bandage contact lenses are widely fitted, and are the ones approved by the FDA for prolonged use after PRK. Currently, a variety of contact lenses are used as therapeutic soft contact lenses after PRK like Lotrafilcon A (Focus Night & Day, Ciba Vision), Lotrafilcon B (O 2 Optix, Ciba Vision), Senofilcon A (Acuvue Oasys, Vistakon Inc.), Balafilcon A, Omafilcon A (Proclear, Cooper Vision) and Senofilcon A. Lotrafilcon B is approved by FDA for 6 days of continuous wear and Senofilcon A for 1 week of continuous wear, while Lotrafilcon A...
Corticosteroids and Non-steroidal Anti-inflammatory Agents (NSAIDs) Therapy

It is necessary to distinguish between corneal haze that appears in the first weeks or months after PRK and pathologi- cal corneal haze that appears as a result of myofibroblasts. If the corneal haze persists over time, it may cause a corneal opacity and the thickening of the tissue that would result in a regression of the refractive error, decreased visual acuity and irregular astigmatism. Clinically significant corneal haze occurs in 0.5%-5% of the cases. Corneal haze that most commonly occurs after PRK is not clinically significant, and is not attributed to myofibroblasts. According to Wilson, in human corneas that develop late corneal haze after PRK, the resolution of the opacity is slow, and the restatement of the refractive correction is produced between 1 and 3 years postoperatively. It has been postulated that the extinction of corneal haze can be influenced by the disappearance of myofibroblasts, reabsorption of abnormal extracellular matrix (ECM) and restoration of normal corneal structure.

After surgery, a variety of drugs are prescribed to avoid corneal haze, for instance, corticosteroids – antiinflammatories to avoid the pain and inflammation-, plasmin inhibitors, growth factors or antimetabolites. Topical therapy after PRK prevents complications like keratitis, infections or corneal haze. The most common treatment after PRK to avoid the corneal inflammation is the application of corticosteroids. Corticosteroids are not recommended for long periods because of their side effects, like intraocular pressure (IOP) rise and the risk of cataracts. Corticosteroids delay epithelial healing. When corneal haze appears 2–3 months after PRK, the clinical observations confirm that haze is "corticosteroid-responsive" in 10%-15% of patients. Researchers disagree about the benefit of corticosteroids to reduce the corneal haze after PRK. According to Wilson, the topical administration of 1% prednisolone acetate (Pred Forte) quickly removes the corneal opacity and produces a change in refractive error. In the remaining 85% or 90% of cases, the corticosteroids do not exert any change. Corticosteroids could be replaced by non-steroidal anti-inflammatory agents (NSAIDs), tramistat, cysteine or antioxidants like Vitamine E. NSAIDs are effective in reducing corneal pain, postoperative photophobia and inflammation. The inflammatory response is mediated by prostaglandins synthesized from arachidonic acid by cyclooxygenase 1 (COX-1) or cyclooxygenase 2 (COX-2). The antiinflammatory and analgesic properties of the nonsteroidal anti-inflammatory drugs (NSAIDs) are achieved by the inhibition of COXs activity. The use of certain steroidal and non-steroidal anti-inflammatory drugs (NSAID) delay reepithelialization and increase the risk of haze formation, although the results are still contradictory. Vetrugno et al. proved that 0.1% fluoromethalone acetate administered in the first day after PRK reduced corneal haze and myopic regression, particularly in high myopic patients. NSAIDs like diclofenac and ketorolac have shown reduction in the pain sensation, but also a significant delay in corneal reepithelialization after PRK. Nepafenac (Nevanac; Alcon Laboratories Inc., Ft Worth, Tex) is a new topical NSAID with greater corneal permeability that has been approved for the treatment of inflammation after surgery. Jalali et al. found that 0.1% Nepafenac did not increase haze formation, neither hamper corneal epithelial healing, but they did.
not found statistically significant differences in corneal reepithelization between nepafenac and non-nepafenac groups (P=.61). Caldwell et al. in a randomized double-masked study, demonstrated that 0.1% nepafenac was safe for corneal reepithelialization, and reduced the post-operative pain at day 1 (0.76 versus 1.68) and day 2 (1.26 versus 2.23) compared with the placebo group (P<.0005). Other NSAIDs for corneal pain reduction are also available, like Bromfenac, Flurbiprofen sodium and Indomethacin.\(^9\)

Despite the presence of complications is low, non-steroidal anti-inflammatory drugs (NSAIDs) may produce conjunctival hyperemia, transient burning, stinging, superficial punctate keratitis, epithelial defects, subepithelial infiltrates, corneal melting and perforation.\(^7,17\) However, Caldwell et al.,\(^8,9\) in a randomized double-masked study, proved that 0.1% nepafenac did not have adverse effects. Postoperative oral analgesics, like NSAIDs, are able to produce gastrointestinal, cardiovascular, respiratory and central nervous system complications.\(^17,18\) Another treatment that is widespread for the inhibition of inflammation and for treatment of dry eye is the Cyclosporine A, with doses from 0.05% to 2.00%.\(^9\) Nien et al.\(^9\) used Cyclosporine A 0.05% and prednisolone acetate 0.01% to compare the effect in corneal haze prevention in rabbit corneas following PRK. They concluded that Cyclosporine A did not have any effect, whereas prednisolone acetate was effective in reducing short-term corneal haze, but did not prevent corneal fibrosis.\(^9\)

**Alternative Therapies for Corneal Haze Prevention**

The use of drugs does not completely suppress corneal haze formation after PRK. Research has focused on new therapies that could prevent corneal haze, like genetic evaluation of type IV collagens synthesis.\(^23\) Lumican and keratocan genes have also been evaluated for management of subepithelial persistent corneal haze after PRK, but without a consistent finding.\(^183\) It has been postulated that vitamin E, probucol or heparin may inhibit collagen type IV synthesis, but they have not been approved for topical use because of their adverse effects.\(^23\) Vitamin E and amino acids play an important role in corneal reepithelialization and in the prevention of corneal haze and keratocyte apoptosis, especially in high myopia.\(^2,77\) A preliminary clinical trial concluded that oral supplementation with vitamin A and vitamin E accelerated the reepithelialization, and reduced corneal haze formation,\(^183\) but it seems that the topical administration of vitamin A alone do not have any effect.\(^184\) Alternative treatments to MMC that prevent corneal haze formation, but produce less damage to keratocyte are bevacizumab and ramapycin.\(^185\) Subconjunctival injection of PRM-151 could presumably prevent corneal haze, as it inhibits the pro-fibrotic myofibroblast differentiation.\(^186\) Trichostatin A, similarly, prevents myofibroblast formation by inhibiting TGF-beta1.\(^187\) As cytokines and growth factors control the synthesis of collagen type IV, they might be also useful treatments for corneal haze prevention.\(^185\) PRK increases the release of leukocytes, TGF-β1, TNF-α, and PDGF-BB in human tears during the first days of wound healing.\(^185,30\) TGF is a cytokine released by the lacrimal gland, corneal epithelium and conjunctival cells.\(^179\) Three forms of TGF-β exist (TGF-β1, TGF-β2 and TGF-β3) and each one is involved in the wound healing process in a different way. TGF-β1 is increased in early epithelial healing, and exerts an influence in the subepithelial fibrosis formation and activation of keratocytes after PRK.\(^12\) Bühren et al.\(^179\) proved that the application of anti-TGF-β in felines reduced the differentiation in vitro of keratocytes into myofibroblast, and corneal haze diminished. They suggested that this reduction in differentiation improved optical quality. The combination of the nerve growth factor (NGF) and decosahexaenoic acid stimulated the regeneration of basal epithelial cells in rabbits after PRK,\(^12\) which is imperative for a proper wound healing. Medduri et al.\(^35\) studied the effect of basic fibroblast growth factor (b-FGF) in circumstances of delayed healing after PRK.\(^9\) 50 patients were enrolled in b-FGF eye drop treatment group and 50 patients in saline drops (placebo) group. They observed greater corneal epithelial healing in the b-FGF group than in the placebo group at 4 days (98% versus 72%, respectively) and 5 days after surgery (100% versus 92%, respectively).\(^35\)

Artificial tears are the most widely used solution for corneal lubrication. However, they do not have biological components that promote corneal regeneration. In fact, they contain stabilizers, preservatives, or other additives that may induce toxic or allergic reactions.\(^21\) Blood derivates as plasma rich in growth factors are an alternative to artificial tears, and have not possibility of rejection.\(^31\) Anitua et al.\(^51\) proved that plasma rich in growth factors obtained from patient’s blood enhanced corneal healing, and reduced the formation of corneal haze. The difference between the plasma rich in growth factors group (PRGF-Endoret treatment) and control group was negligible at day 3. They attributed it to the increase of proliferative cells (Ki-67\(^+\)) in the control group. They suggested that the increase in proliferative cells could be associated with epithelial hyperplasia observed at day 3 and 7 after PRK in control group.\(^51\) They also found that the epithelium of the PRGF-Endoret group was formed by 5–6 layers.\(^51\)

**Corneal Nerve Regeneration and Neuropathic Corneal Pain Management**

The regeneration of the corneal nerves after PRK is associated with the improvement of cellular integrity.\(^22\) To date, few therapeutic treatments have been developed for nerve regeneration. Javaly et al.\(^188\) investigated the benefits of topical platelet-rich plasma, but subbasal nerve density did not improve after 3 months of treatment compared to controls (P=.66). Studies in animal models have demonstrated more encouraging results. Esquenazi et al.\(^25\) studied the outcomes of the combination of nerve growth factor (NGF) and docosahexaenoic acid (DHA) in rabbits in promoting corneal nerve regeneration. They observed that this combination increased corneal nerve regeneration, as well as epithelial proliferation and decreased rose bengal staining compared to the application of NGF or DHA alone. Cortina et al.\(^188\) showed similar results with pigment epithelial-derived factor (PEDF) plus docosahexaenoic acid (DHA).
Moreover, this combination proved to enhance corneal sensitivity. Recent evidence suggests that peripheral nerve system regeneration and inflammatory processes share common pathways, and some degree of inflammation is required for neuroregeneration. Therefore, cyclosporine A and corticosteroid treatments could interfere in a proper nervous recovery.

When corneal nerve regeneration process fails, corneal neuropathic pain might take place. The up- and down-regulation of ion channels in axotomized nerves can change the excitability of fibers, and produce spontaneous discharges and altered sensitivity to exogenous stimuli. This would result in corneal pain non-treatable with aforementioned drugs. Anticonvulsants, opiates and topical local anesthetics can manage corneal neuropathic pain. The anticonvulsant Gabapentin (Neurontin) is an analog of gamma-aminobutyric acid (GABA), and its reliability in treating corneal pain is conflicting, mainly because of a lack of studies. Lichtinger et al. compared in a prospective randomized, double-blind, placebo-controlled study the efficacy of Gabapentin in the reduction of the corneal pain. They administered gabapentin capsules (300 mg) in 20 patients and additional 20 patients received identical placebo capsules. They demonstrated that gabapentin reduced corneal pain during the first 24 h (P=.003), at postoperative day 1 (P=.002), between 24 and 48 h (P=.024), at postoperative day 2 (P=.018) and between 48 and 72 h (P=.001). Farkorvich et al. tried to prove the efficacy of topical opioid in the treatment of pain, concluded in a double-blind randomized prospective study that the administration of 0.5% morphine drops was an effective and safe method to control of post-PRK pain, and did not hamper epithelial healing or refractive outcomes. Topical local anesthetics include tetracaine, proparacaine, lidocaine and bupivacaine can be also used. Topical tetracaine has been documented to be successful in pain control management and does not produce delayed corneal healing times. However, it produces keratocyte toxicity and keratitis. Topical anesthetics should be used cautiously and for short-term treatments. Antidepressants are prescribed for neuropathic pain management elsewhere in the body. To date, no study has been published evaluating the effect of antidepressants for treating post-PRK corneal pain.

Conclusions

Photorefractive keratotomy disrupts corneal structure affecting epithelium, Bowman’s membrane, and anterior stroma. Corneal nerves are severed, which alters corneal integrity and function temporarily. The subsequent corneal wound healing is a complex process that is regulated by a variety of factors. A balance of peptides will determine the final outcome, and the presence of postoperative complications. Corneal wound healing process can be managed with several drugs to enhance regeneration and prevent corneal haze and pain after PRK. Further research in this field is required to completely understand post-PRK corneal regeneration in order to prevent complications, and provide outstanding visual outcomes.

References


44. Martinez-Garcia MC, Merayo-Lloves J, Blanco-Mezquita T, Mar-Sardana S. Wound healing following
49. Efron N. Contact lens-induced changes in the anterior eye as observed in vivo with the confocal microscope. Prog Retin Eye Res. 2007;26:398–436.


